ORGANIC LETTERS

2011
Vol. 13, No. 3
498−501

Characterization of the *lnmKLM* Genes Unveiling Key Intermediates for β-Alkylation in Leinamycin Biosynthesis

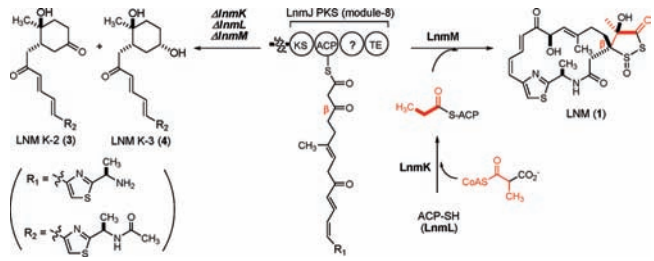
Yong Huang,[†] Sheng-Xiong Huang,[†] Jianhua Ju,[†] Gongli Tang,[†] Tao Liu,[†] and Ben Shen[*,†,‡,§]

Division of Pharmaceutical Sciences, University of Wisconsin National Cooperative Drug Discovery Group, and Department of Chemistry, University of Wisconsin, Madison, Wisconsin 53705-2222, United States

bshen@pharmacy.wisc.edu

Received November 23, 2010

ABSTRACT



Leinamycin (LNM, 1) biosynthesis is proposed to involve β-alkylation of the polyketide intermediate, catalyzed by LnmKLM. Inactivation of *lnmK*, *lnmL*, or *lnmM* afforded mutant strains that accumulated LNM K-1 (2), K-2 (3), K-3 (4), and isomers LNM K-1′ (5), K-2′ (6), and K-3′ (7) whose polyketide origin was established by feeding experiments with sodium [1-^{13}C]acetate. These findings confirm the indispensability of LnmKLM in 1 biosynthesis and suggest that β-alkylation proceeds on the growing polyketide intermediate while bound to the LNM polyketide synthase.

β-Alkylations contribute to the vast structural diversity displayed by polyketide natural products, with the β-alkyl branches often resulting from the activity of hydroxymethylglutaryl-CoA (HMG-CoA) synthase homologues (HCSs).[1] HCS catalyzes condensation of acyl-*S*-acyl carrier protein (ACP) with the β-carbonyl group of the polyketide synthase (PKS)-ACP-tethered growing polyketide intermediate to afford a β-hydroxymethylacyl-*S*-ACP intermediate, which can undergo further dehydration, decarboxylation, or both by two enoyl-CoA hydratase homologues (ECH1 and ECH2) to afford a β-alkylated intermediate.[1] This pathway for β-methyl branch installation has been extensively studied, and a dedicated set of three proteins has been identified that derives acetyl-*S*-ACP from malonyl-CoA.[1−4]

Leinamycin (LNM, **1**), a potent antitumor antibiotic, possesses a β-branched C3 unit that is part of the unique five-membered 1,3-dioxo-1,2-dithiolane moiety.[5] We previously cloned, sequenced, and characterized the *lnm* biosynthetic gene cluster from *Streptomyces atroolivaceus* S-140.[6] These studies established that the 18-membered macrolactam backbone of **1** is assembled by a hybrid nonribosomal peptide

[†] Division of Pharmaceutical Sciences.
[‡] University of Wisconsin National Cooperative Drug Discovery Group.
[§] Department of Chemistry.

(1) Calderone, C. T. *Nat. Prod. Rep.* **2008**, *25*, 845–853.
(2) Calderone, C. T.; Kowtoniuk, W. E.; Kelleher, N. L.; Walsh, C. T.; Dorrestein, P. C. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, *103*, 8977–8982.
(3) (a) Gu, L.; Jia, J.; Liu, H.; Hakansson, K.; Gerwick, W. H.; Sherman, D. H. *J. Am. Chem. Soc.* **2006**, *128*, 9014–9015. (b) Buchholz, T. J.; Rath, C. M.; Lopanik, N. B.; Gardner, N. P.; Hakansson, K.; Sherman, D. H. *Chem. Biol.* **2010**, *17*, 1092–1100.
(4) (a) Simunovic, V.; Müller, R. *ChemBioChem* **2007**, *8*, 497–500. (b) Simunovic, V.; Müller, R. *ChemBioChem* **2007**, *8*, 1273–1280. (c) Calderone, C. T.; Iwig, D. F.; Dorrestein, P. C.; Kelleher, N. L.; Walsh, C. T. *Chem. Biol.* **2007**, *14*, 835–846.
(5) (a) Hara, M.; Asano, K.; Kawamoto, I.; Takiguchi, T.; Katsumata, S.; Takahashi, K. I.; Nakano, H. *J. Antibiot.* **1989**, *42*, 1768–1774. (b) Hara, M.; Takahashi, I.; Yoshida, M.; Asano, K.; Kawamoto, I.; Morimoto, M.; Nakano, H. *J. Antibiot.* **1989**, *42*, 333–335.

10.1021/ol102838y
Published on Web 12/30/2010

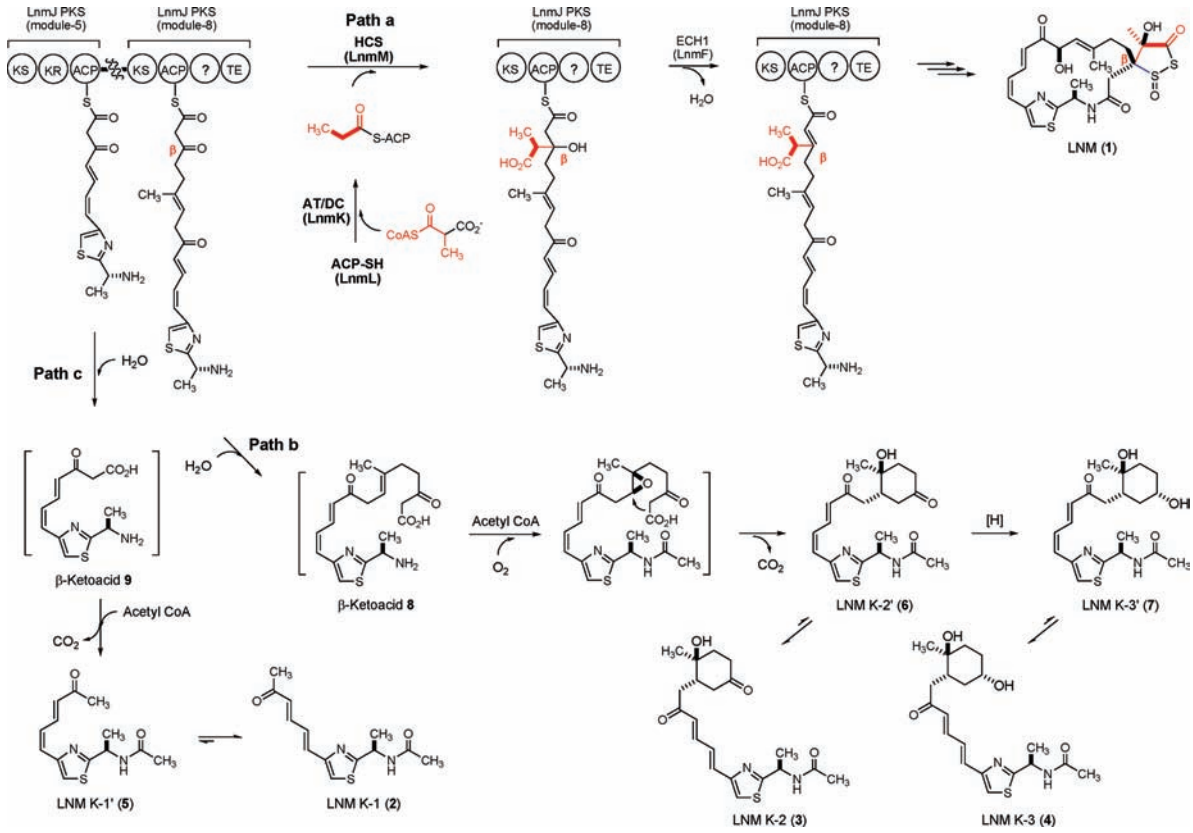


Figure 1. Proposed LNM (**1**) biosynthetic pathway featuring LnmKLM-catalyzed β-alkylation of the LnmJ PKS module-8 ACP-tethered intermediate in the *S. atroolivaceus* S-140 wild-type strain (path a) and processive elongation of polyketide intermediates tethered to the LnmJ PKS module-5 and module-8 ACPs as evidenced by accumulation of LNM K-1 (**2**), K-2 (**3**), K-3 (**4**), K-1′ (**5**), K-2′ (**6**), and K-3′ (**7**) in the *S. atroolivaceus* S-140 mutant strains SB3029 (i.e., Δ*lnmK*), SB3030 (i.e., Δ*lnmL*), and SB3031 (i.e., Δ*lnmM*) (paths b and c).

synthetase (NRPS)-PKS and suggested that the β-branched C3 unit is installed by a set of four proteins: LnmL, an ACP; LnmK, a bifunctional acyltransferase/decarboxylase (AT/DC); LnmM, an HCS homologue; and LnmF, an ECH1 homologue. It has been suggested that LnmM catalyzes condensation of propionyl-*S*-LnmL with the LnmJ PKS module-8 ACP-tethered polyketide intermediate, followed by LnmF-catalyzed dehydration, to afford the β-branched C3 unit by the same mechanism as that used for β-methyl branch incorporation with the exception of a missing decarboxylation step catalyzed by ECH2.[7] However, the origin of propionyl-*S*-ACP is distinct from that of acetyl-*S*-ACP.[7] The bifunctional AT/DC activity of LnmK has been confirmed to derive propionyl-*S*-LnmL from methylmalonyl-CoA by acylation of LnmL and subsequent decarboxylation (Figure 1).

We now report *in vivo* characterization of the *lnmKLM* genes to support their assigned roles in the installation of the β-branched C3 unit for **1** biosynthesis. Inactivation of *lnmKLM* in *S. atroolivaceus* S-140 afforded mutant strains SB3029, SB3030, and SB3031. Each of the mutant strains failed to produce **1** but accumulated the same set of six new metabolites (Figure 2). Isolation and structural elucidation of the six new metabolites LNM K-1 (**2**), K-2 (**3**), K-3 (**4**), K-1′ (**5**), K-2′ (**6**), and K-3′ (**7**) support the previously proposed pathway[6,7] and unveil new intermediates for the biosynthesis of **1** (Figure 1).

To explore the roles of LnmK, LnmL, and LnmM *in vivo*, inactivation of *lnmK*, *lnmL*, and *lnmM* was accomplished by inserting an apramycin resistance gene cassette into the individual gene within the *lnm* gene cluster in the *S. atroolivaceus* S-140 wild-type.[8] The resultant mutant strains were named SB3029 (i.e., Δ*lnmK*), SB3030 (i.e., Δ*lnmL*), and SB3031 (i.e., Δ*lnmM*), whose genotypes were confirmed by Southern analysis (Figure S1, Supporting Information).

S. atroolivaceus S-140 mutant strains SB3029, SB3030, and SB3031 were fermented, with the wild-type as a control, and their culture supernatants were extracted with ethyl acetate and extracts were analyzed by HPLC.[8] Relative to wild-type, all three mutants were unable to produce **1**, confirming the indispensability of LnmK, LnmL, and LnmM in **1** biosynthesis. Instead, a series of new peaks (**2**−**7**) with

(6) (a) Cheng, Y.; Tang, G.; Shen, B. *J. Bacteriol.* **2002**, *184*, 7013–7024. (b) Cheng, Y.; Tang, G.; Shen, B. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 3149–3154. (c) Tang, G.; Cheng, Y.; Shen, B. *Chem. Biol.* **2004**, *11*, 33–45. (d) Cheng, Y.; Tang, G.; Shen, B. *J. Nat. Prod.* **2006**, *69*, 387–393. (e) Tang, G.; Cheng, Y.; Shen, B. *J. Biol. Chem.* **2007**, *282*, 20273–20282.

(7) Liu, T.; Huang, Y.; Shen, B. *J. Am. Chem. Soc.* **2009**, *131*, 6900–6901.

(8) See Supporting Information for experimental details.

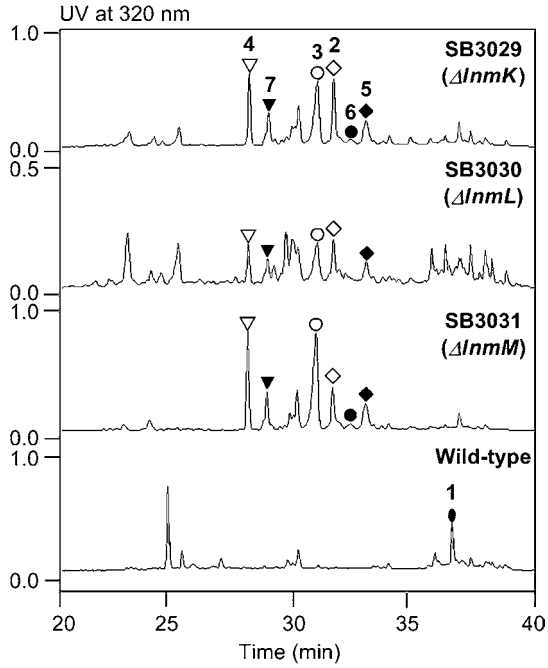


Figure 2. Metabolite profiles of *S. atroolivaceus* S-140 mutant strains SB3029 (i.e., Δ*lnmK*), SB3030 (i.e., Δ*lnmL*), and SB3031 (i.e., Δ*lnmM*), in comparison with the wild-type, upon HPLC analysis. See Figure 1 for structures of **1**−**7**.

retention times shorter than that for **1** were observed upon HPLC analysis; all compounds showed a maximum UV absorption at 320 nm, characteristic of the conjugated (*Z*,*E*)-thiazol-5-yl-penta-2,4-dienone moiety of **1** (Figure 2). LC-ESI-MS analysis revealed that **2**−**7** in the three mutant strains SB3029, SB3030, and SB3031 afforded the same set of $[M + H]^+$ ions at *m*/*z* of 391, 393, 265, 391, 393, and 265, respectively, suggesting that the three mutant strains accumulated the same series of metabolites. The Δ*lnmK* mutant strain SB3029 was then cultured under standard conditions, and **2**−**7** were purified by silica gel column chromatography and semipreparative C-18 reversed phase HPLC; their structures were solved by MS and ^{1}H and ^{13}C NMR spectroscopy.[8]

The molecular formula of **4** was determined to be $C_{20}H_{28}N_2O_4S$ by high resolution MALDI MS measurement, affording an $[M + Na]^+$ ion at *m*/*z* 415.1653 (calculated $[M + Na]^+$ ion at *m*/*z* 415.1667). A comparison of the ^{1}H and ^{13}C NMR spectra of **4** with those of **1** revealed that signals in the range of C-9 to C-17 of **1** were similarly present in **4**, except for the lack of one H (H-11 at δ_H 8.03) in **1** and the presence of one additional H (H-11 at δ_H 7.32) in **4**. The existence of the conjugated (*E*,*E*)-double bond in the region of C-10 to C-13 was supported by the presence of their unique coupling constants ($J > 14.0$ Hz), as well as gCOSY and HMBC correlations (Figure 3). The remaining proton and carbon signals suggested the existence of a cyclohexane moiety, which was assigned on the basis of one H−H coupling system (H-7, -2, -3, -4, and -5) and HMBC correlations between H_3-18 (δ_H 1.01) and C-5 (δ_c 40.1), C-6

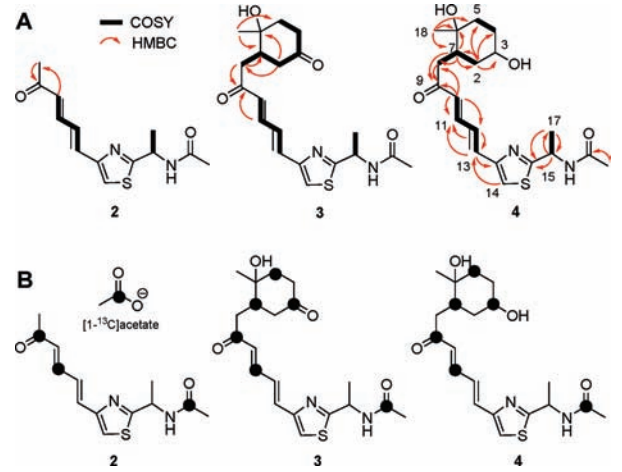


Figure 3. (A) Key HMBC and COSY correlations for LNM K-1 (**2**), K-2 (**3**), and K-3 (**4**). Similar HMBC correlations of **2**, **3** with **4** were omitted for clarity. (B) Specific ^{13}C-enrichment of **2**, **3**, and **4** resulting from feeding experiments with sodium [1-^{13}C]acetate.

(δ_c 70.1), and C-7 (δ_c 41.8) (Table S2, Figures S9, S10, Supporting Information). The relative stereochemistry of C-3, C-6, and C-7 was established on the basis of ROESY correlations of H-3 with H-7 and of H_3-18 with H-8, while the absolute stereochemistry of C-16 was deduced on the basis that **1** and **4** share the same biosynthetic origins (Figure 1). The structure of **4** was fully consistent with other correlations as observed in gCOSY, gHMBC, and HMQC.

The structure of **3** was established in a fashion analogous to **4** by ^{1}H and ^{13}C NMR experiments. The molecular formula of **3** was determined to be $C_{20}H_{26}N_2O_4S$ on the basis of high resolution MALDI MS measurement, affording an $[M + Na]^+$ ion at *m*/*z* 413.1480 (calculated $[M + Na]^+$ ion at *m*/*z* 413.1505). In comparison to **4**, **3** lacks the oxygenated methine signal (C-3 at δ_C 68.6 for **4**) and has a characteristic ketone carbonyl carbon (C-3 at δ_C 210.3 for **3**) (Table S2, Figures S5, S6, Supporting Information), in agreement with its molecular weight being 2 Da lower than that of **4**.

Compounds **2** and **5** cannot be completely separated from each other due to their rapid interconversion.[8] They both yielded $[M + H]^+$ ions at *m*/*z* 265 upon LC-ESI-MS analysis, suggesting the existence of a pair of regioisomers. The molecular formula of **2** and **5** was determined to be $C_{13}H_{16}N_2O_2S$ by high resolution MALDI MS measurement, yielding an $[M + H]^+$ ion at *m*/*z* 265.1001 (calculated $[M + H]^+$ ion at *m*/*z* 265.1005), and their structures were assigned on the basis of ^{1}H and ^{13}C NMR spectra of the mixture of **2** and **5** (Table S2, Figures S3, S4, Supporting Information). To confirm the interconversion of **2** and **5**, a mixture of both substances was exposed to UV light (254 nm) for a period of 30 min in $CDCl_3$,[8] and ^{1}H NMR spectra before and after irradiation were compared (Figure S2, Supporting Information). Diminished proton signals of **5** corresponded to the compound's conversion to **2**, verifying that **5** is the photoisomer of **2**; **2** is the more photostable conformation, consistent with the proposed structures (Figure 1).

Similar to **2** and **5**, compounds **6** and **7** cannot be fully separated from **3** and **4**, respectively, due to the rapid conversion of **6** to **3** and **7** to **4**. On the basis of LC-ESI-MS analyses, which afforded the same $[M + H]^+$ ions at *m*/*z* 391 for **3** and **6** and at *m*/*z* 393 for **4** and **7**, **3**/**6** and **4**/**7** were each assigned to be a pair of photoisomers (Figure 1) with the structure of **6** further confirmed by ^1H and ^{13}C NMR spectra of the **3**/**6** mixture (Table S2, Figures S7, S8, Supporting Information).

The structures of **2**−**7** differ from **1** most notably by virtue of the absence of the 18-membered macrolactam, acetylation of the D-alanine N-terminus, and the cyclohexane moieties in **3**, **4**, **6**, and **7**. To determine their biosynthetic origin and to correlate their intermediacy in **1** biosynthesis, sodium [1-^{13}C]acetate was fed to the Δ*lnmK* mutant strain SB3029 and ^{13}C-enriched **3** and **4**, as well as the mixture of **2** and **5**, were isolated and characterized (Table S3, Figures S11−S14, Supporting Information). Thus, as depicted in Figure 3B, specific ^{13}C-enrichments at C-3, C-5, C-7, C-9, and C-11 in **3** and **4** and at C-9 and C-11 in **2** were consistent with the hypothesis that all were biosynthesized by the LNM NRPS-PKS en route to **1** (Figure 1). The C-2 carbon in **3** and **4** was not labeled, suggesting that it originated from C-2 of acetate with the loss of the ^{13}C-labeled C-1 carbon. Similarly, the C-8 carbon in **2** was not labeled, indicating that it was also derived from the C-2 of acetate (Figures 1 and 3).

In this study, we have confirmed that *lnmK*, *lnmL*, and *lnmM* are essential for **1** biosynthesis, experimentally supporting their predicted roles in β-alkylation steps for **1**. These findings shed new light into the initial steps leading to the formation of the dithiolane ring (Figure 1). In brief, following the completion of the final chain extension, the HCS homologue LnmM catalyzes Claisen condensation between the LnmJ PKS module-8 ACP-tethered polyketide intermediate and the propionyl-*S*-LnmL. This results in the attachment of the propionyl group, as a β-alkyl branch, to the ACP-tethered intermediate, which is subsequently dehydrated by the ECH1 homologue LnmF and further modified en route to **1** (Figure 1, path a). Without the β-alkylation steps by LnmKLM, the LnmJ PKS module-8 ACP-bound polyketide intermediate apparently cannot be cyclized by the LnmJ-thioesterase (TE) domain, thereby undergoing hydrolytic release to afford the linear intermediate **8** (Figure 1, path b). This reaction could be catalyzed by either the LnmJ-TE domain, a type I TE, or LnmN, a type II TE within the *lnm* gene cluster.[6] Hydrolytic removal of stalled polyketide or peptide intermediates from PKS or NRPS machinery by both type I and type II TEs is well-known in polyketide, peptide, and hybrid polyketide-peptide biosynthesis.[9] Finally, the β-ketoacid **8** could undergo a series of transformations to afford **3**, **4**, **6**, and **7**, and these reactions are most likely catalyzed by promiscuous enzymes residing outside of the *lnm* cluster (Figure 1, path b).

Interestingly, the mutant strains SB3029, SB3030, and SB3031 also accumulated **2** and **5**. Similar phenotypes have also been observed in mupirocin biosynthesis, where inactivation of the genes encoding for β-methyl incorporation produced shortened shunt metabolites.[10] Thus, **2** and **5** are most likely shunt metabolites of the proposed β-ketoacid **9**, which undergoes release from LnmJ PKS module-5 in a mechanism similar to the aforementioned (Figure 1, path c). This result suggests the importance of specific protein−protein interactions among LnmKLM and LnmJ PKS, the disruption of which leads to the premature release of LnmJ PKS ACP-tethered polyketide intermediates, derailing **1** biosynthesis (Figure 1). The above hypothesis was further supported by the observation that the start and stop codons between *lnmJ*, *lnmK*, *lnmL*, and *lnmM* overlap, suggesting the cotranslation of these genes, thereby ensuring the formation of a proper protein complex.

The accumulation of **2**−**7** in mutant strains SB3029, SB3030, and SB3031 supports a processive mechanism for the LNM NRPS-PKS megasynthase-catalyzed assembly of the LNM hybrid peptide-polyketide backbone from amino acid and short carboxylic acid precursors. Although PKS or hybrid PKS-NRPS systems are widely accepted to assemble polyketides or hybrid peptide-polyketides processively, this has been demonstrated experimentally in only a few cases. For example, inactivation of the amide transferase gene *rifF* in *Amycolatopsis mediterranei* abolished rifamycin production, giving rise instead to a series of linear polyketides with chain lengths varying from tetra- to decaketide.[11] In bleomycin[12] and epothilone[13] biosynthesis, various peptides or polyketides representative of varying elongation steps have also been identified. Predicated on these findings and those reported here, the processivity of PKS, NRPS, or hybrid PKS-NRPS represents a promising tool by which to generate natural product structural diversity by rational manipulation of these processive biosynthetic machineries.

Acknowledgment. We thank Kyowa Hakko Kogyo Co. Ltd. (Tokyo, Japan) for the wild-type *S. atroolivaceus* S-140 strain and the Analytical Instrumentation Center of the School of Pharmacy, UW-Madison, for support in obtaining MS and NMR data. This work was supported in part by NIH Grants CA106150 and CA113297.

Supporting Information Available: Available detailed experimental procedures, ^1H and ^{13}C NMR spectra for compounds **2**−**7**. This material is available free of charge via the Internet at http://pubs.acs.org.

OL102838Y

(9) (a) Sieber, S. A.; Marahiel, M. A. *Chem. Rev.* **2005**, *105*, 715–738. (b) Du, L.; Lou, L. *Nat. Prod. Rep.* **2010**, *27*, 255–278.

(10) Wu, J.; Hothersall, J.; Mazzetti, C.; O'Connell, Y.; Shields, J. A.; Rahman, A. S.; Cox, R. J.; Crosby, J.; Simpson, T. J.; Thomas, C. M.; Willis, C. L. *ChemBioChem* **2008**, *9*, 1500–1508.

(11) Yu, T. W.; Shen, Y. M.; Doi-Katayama, Y.; Tang, L.; Park, C.; Moore, B. S.; Hutchinson, C. R.; Floss, H. G. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96*, 9051–9056.

(12) (a) Takita, T.; Muroka, Y. *Biosynthesis and chemical synthesis of bleomycin*; Walter de Gruyter: New York, 1990; pp 289−309. (b) Du, L.; Sanchez, C.; Chen, M.; Edwards, D. J.; Shen, B. *Chem. Biol.* **2000**, *7*, 623–642.

(13) Hardt, I. H.; Steinmetz, H.; Gerth, K.; Sasse, F.; Reichenbach, H.; Hofle, G. *J. Nat. Prod.* **2001**, *64*, 847–856.